MAYO CLINIC

LEVERAGING METHYLATED DNA MARKERS IN THE DETECTION OF OVARIAN AND ENDOMETRIAL CANCERS

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DISCLOSURE OF RELEVANT FINANCIAL RELATIONSHIP(S) WITH INELIGIBLE COMPANIES

 Dr. Bakkum-Gamez is listed as an inventor on intellectual property jointly owned by Mayo Clinic and Exact Sciences.

REFERENCES TO OFF-LABEL USAGE(S) OF PHARMACEUTICALS OR INSTRUMENTS

- Data presented is research-only.
- No references to off-label usage of pharmaceuticals or instruments.

OBJECTIVES

- Overall: Focus on bench-to-bedside cancer detection biomarker development in endometrial cancer (EC) and ovarian cancer (OC)
- Current clinical state in EC diagnostics
- Sampling of EC biomarker work
- Mayo Clinic methylated DNA marker pipeline in EC and OC
- Ongoing work and future directions

THE NUMBERS – POSTMENOPAUSAL BLEEDING AND ABNORMAL UTERINE BLEEDING

• Postmenopausal bleeding (PMB) chief complaint 5% office GYN evaluations

• PMB: affects 4-11% women in their lifetime

10% with PMB have EC

50M postmenopausal women in the US

• Abnormal uterine bleeding (AUB) chief complaint for 70% of *peri-*/postmenopausal GYN outpatient evaluations

- AUB: affects 1.4M women/year in US age 18-50
 - Perimenopause: is bleeding change normal or abnormal?

5% with perimenopausal AUB have EC

Cozza, et al. Eur. Rev. Med. Pharmacol Sci. 2017. Moodley and Roberts, J Obstet Gynaecol. 2004.

WORK UP FOR AUB OR PMB

- Any of the following:
 - Transvaginal ultrasound
 - Office hysteroscopy
 - Sonohysterogram
 - Endometrial sampling criterion standard*
 - Office-based biopsy
 - D&C



*ACOG Guidelines → All women ≥45 yo, endometrial sampling should be performed as first-line test Plus: Endometrial sampling should also be performed in women <45 yo with obesity or other EC risk factors

 Hysterectomy may be needed for diagnostic purposes if benign etiology cannot be ruled out
 ACOG Cmte Opinion, Number 557, Reaffirmed 2020.

> Munro, et al. Int J Gynecol Obstet. 2011 ©2025 Mayo Foundation for Medical Education and Research | slide-5

CHALLENGES WITH CURRENT DIAGNOSTIC APPROACH

- Patient needs to see a provider to perform <u>every</u> test
 - Access
 - Lay population awareness of symptoms
 - Diversity in patient perception of AUB
- Prior benign-pathology therapies can preclude successful endometrial sampling
 - For example: Endometrial ablation (40% fail endometrial sampling)
 - Endometrium found to be 100% present in post-ablation hysterectomies
 - Major surgery to dx small % EC

PATIENTS EXPERIENCE PAIN WITH OFFICE BIOPSY

Visual Analog Scale (VAS) pain scores among **190 patients** with VAS scoring available for all three sampling methods (EMB, Tao brush, tampon)



Bagaria, et al. Gyn Onc. 2021.

LEVERAGING CARCINOGENESIS IN BIOMARKER DEVELOPMENT

• What molecular changes happen between benign and cancer?

- DNA mutations, epigenetic changes, CNVs
- Microorganism genomes

Altered transcription and translation

- Can we use these changes as signals to detect precancerous and early stage EC?
- Can we detect those changes in minimally-invasively collected biospecimens?
- Can we leverage self-collected biospecimens?

COLOGUARD™ MULTI-TARGET STOOL DNA TEST: A STORY OF SELF-SAMPLING SUCCESS

- Simple device for collection & mailing
- Preservative buffer
- Targets multiple markers
 - Methylated BMP3 & NDRG4
 - Mutant KRAS
 - *β-actin* (human DNA)
 - Hemoglobin (FIT)
- Sensitive multiplex DNA assay (QuARTS)
- Screening in average risk population; not a diagnostic test
- FDA-Approved: August 2014

Slide courtesy of John Kisiel, MD. Mayo Clinic Gastroenterology.





A NOVEL LIQUID BIOPSY: VAGINAL FLUID



Fiegl, et al. Ca Epi Bio Prev. 2004. Finan, et al. Gyn Onc (SGO Abstracts). 2012. Bakkum-Gamez, et al. Gyn Onc. 2015. ©2025 Mayo Foundation for Medical Education and Research | slide-10

DNA METHYLATION IN VAGINAL TAMPONS: MAYO PROOF-OF-PRINCIPAL

- 38 EC; 28 benign controls ≥45 yrs of age
- Clinically indicated hysterectomy
- 12 genes, 97 CpGs
- Pyrosequencing
 - Tampon DNA
 - Tao brush endometrial cytology DNA
- Consistent methylation across CpGs
- Higher mean methylation in EC v. benign in 9 genes (ADCYAP1, ASCL2, CDH13, HS3ST2, HTR1B, MME, HAAO, HOXA9, and RASSF1)
- Similar results from tampon and direct endometrial brushing





Bakkum-Gamez, et al. Gyn Onc. 2015. ©2025 Mayo Foundation for Medical Education and Research | slide-11

DISCOVERING NOVEL MOLECULAR MARKERS OF GYNECOLOGIC CANCER

Endometrial cancers of 5 predominant histologies

- Grade 1/2 endometrioid
- Grade 3 endometrioid
- Clear cell
- Carcinosarcoma
- Serous

Type II; aggressive histologies

- *Ovarian cancers (4 predominant histologies), cervical cancers (2 histologies)
- Benign endometrial menstrual phases, cervical, vaginal, tubal tissue, endometrial hyperplasias, cervical dysplasia, blood buffy coat
- Methylome sequencing on tissue-derived DNA

ENDOMETRIAL CANCER – 3-STEP FOCUSED PIPELINE



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INDEPENDENT TISSUE VALIDATION AMONG HISTOLOGIES

AILEC	AUC	FC	Clear Cell EC	AUC	Serous EC	AUC	Carcinosarcoma	AUC	Endometrioid E	AUC
EMX2OS-RS	0.95	29.31	CYTH2-FS	1.00	(EMX2OS-RS)	1.00	(EMX2OS-RS)	0.96	(NBPF8-FS)	0.97
NBPF8-FS	0.92	14.57	OBSCN FS	0.99	LRRC41(8188)-FS	0.94	ZNE506-F8	0.94	MAX.chr8.1451	0.94
CYTH2-F8	0.89	16.18	EMX2OS-RS	0.97	DIDO1-FS	0.94	MAX.chr10.226	0.94	EMX2OS-RS	0.93
MAX.chr10.226244	0.89	77.62	SEPT9-ES	0.97	OBSCN-FS	0.91	(NBPF8-FS)	0.93	SFMBT2.894(1)	0.92
MAX.chr8.1451038	0.88	22.51	MAX.chr14.1030	0.97	CYTH2-FS	0.90	ZNF90-RS	0.90	END.JSRP1-FS	0.91
JSRP1-FS	0.87	3.73	EEF1A2-RS	0.95	MAX.chr8.14510382§	0.88	VILL-RS	0.88	OBSCN-FS	0.89
ZNF90-RS	0.87	6.43	MDFI-FS	0.95	LRRC8D-FS	0.87	KANK1-FS	0.87	MAX.chr10.226	0.88
SFMBT2.894(1000	0.86	22.62	SFMBT2.894(10	0.93	VILL-RS	0.87	JSRP1-FS	0.87	MAX.chr8.1451	0.88
OBSCN-FS	0.86	14.72	GDF7-FS	0.93			LRRC8D-FS	0.86	CYTH2-FS	0.87
MPZ-FS	0.85	112.07	MAX.chr8.14510	0.93			MAX.chr8.1451	0.86	ZNF90-RS	0.87
LRRC8D-FS	0.85	17.70	JSRP1-FS	0.92			OBSCN-FS	0.85	MPZ-FS	0.87
DIDO1-FS	0.85	214.16	DIDO1-FS	0.92						
			MPZ-FS	0.91						
			LRRC34-RS	0.91						
			SQSTM1-FS	0.90						
			ZNF90-RS	0.90						
			ZNF323-FS	0.90						
			LRRC8D-FS	0.90						
			VILL-RS	0.90						
			LRRC41(8188)-F	0.90						
			MAX.chr10.2262	0.88						
			NBPF8-FS	0.86						

TRANSLATION FROM DISCOVERY TO CLINICAL PILOT: TESTING APPLICATION



Biological tissue-based validation 33 MDMs





Tampon clinical pilot; independent subjects 29 MDMs

CLINICAL TAMPON PILOT – BUFFER ENHANCEMENT

EMX20S

- 93 women ≥45 yo with benign AUB/PMB
- 100 EC
- Early buffer (PBS only pre 2/14/17) v. current buffer (PBS/EDTA)
- PBS/EDTA buffer: 52 benign; 57 EC
- 29 MDM-panel
- Greater discrimination in PBS/EDTA buffer

TABLE: Distribution of endometrial cancer (EC) histologies and cross-validated sensitivity by methylated DNA marker panel at 95% specificity in PBS/EDTA tampon buffer (N=57 ECs).

EC histology	Endometrioid	Serous	Carcinosarcoma	Clear cell	Mixed
Ν	20	23	9	3	2
Sensitivity at 95%	85%	78%	89%	67%	50%
(95% CI)	(62-97%)	(56-93%)	(52-99%)	(9-99%)	(1-99%)



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Endometrial Cancer

- Novel biomarker discovery: RRBS, frozen tissue (EC and pan-GYN tract) DNA
- Stringent methylated DNA marker (MDM) filtering criteria (ROC, fold-change, control background
- Blinded, biological validation: msPCR, FFPE tissue (EC and pan-GYN tract) DNA
- Testing in vaginal fluid collected via tampon from women with and without EC**: msPCR for 29 top performing MDMs





Bakkum-Gamez

METHYLATED DNA MARKERS (MDMS) IN THE DETECTION OF OVARIAN CANCER





DESIGNING A SUCCESSFUL EARLY DETECTION TEST: REACHING FOR NIRVANA IN OVARIAN CANCER

- 1. High sensitivity to detect small volume, earliest stage disease
- 2. Encompass broad histologies
- Generalizability to average-risk and elevated risk women
- 4. Effective even in settings of altered gynecologic anatomy
- 5. Widely accessible
- 6. Minimally invasive
- 7. Operator independent (automated)
- 8. Reasonable cost
- 9. For effective screening, needs to decrease mortality from EOC



ANATOMIC LIMITATIONS—*IDEAL* CONTINUITY OF STRUCTURES



Caveat: Potential for continuity to be disrupted with clinically indicated procedures.

OVARIAN CANCER – THE 3-STEP FOCUSED PIPELINE

I. Discovery & Techni Archived fresh frozen OC tissue, Pros	cal Validation pective benign FFPE fallo	opian tube tissue	→ Discovery: 526 genes differentially methylated in OC
Ovarian Cancer (n=57)		Benign (n=29)	\rightarrow 54 selected for technical
18 Endometrioid 15 Clear Cell	6 Mucinous 14 F	TE 19 Buffy Coat	validation
ethylome sequencing (RRBS) (Discover	ery) & Methylation speci	fic PCR (Technical	\rightarrow 44 candidate genes AUC>0.90
II. Biological Va Independent archived FFPE OC tissue Ovarian Cancer (n= GS 28 Endometrioid 25 Clear C ion specific PCR	lidation e, Prospective benign FFF 105) Cell 16 Mucinous	PE fallopian tube tissue Benign (n–31) 31 FTE	→ 33 best performing genes in technical validation
III. Testing in P Archived plasma samples Ovarian Cancer (n=91 IGS 4 LGS 8 Endometrioid 4	Plasma) Clear Cell 2 Mucinous	Benign (n=91) 91 Plasma	→ 11 candidate genes/methylated DNA markers (MDMs)
	I. Discovery & Techni Archived fresh frozen OC tissue, Pros Ovarian Cancer (n=57) 18 Endometrioid 15 Clear Cell nethylome sequencing (RRBS) (Discov II. Biological Va Independent archived FFPE OC tissue Ovarian Cancer (n= GS 28 Endometrioid 25 Clear O tion specific PCR III. Testing in F Archived plasma samples Ovarian Cancer (n=91 IGS 4 LGS 8 Endometrioid 4	I. Discovery & Technical Validation Archived fresh frozen OC tissue, Prospective benign FFPE falls Ovarian Cancer (n=57) 18 Endometrioid 15 Clear Cell 6 Mucinous 14 F hethylome sequencing (RRBS) (Discovery) & Methylation speci II. Biological Validation Independent archived FFPE OC tissue, Prospective benign FFF Ovarian Cancer (n=105) GS 28 Endometrioid 25 Clear Cell 16 Mucinous tion specific PCR III. Testing in Plasma Archived plasma samples Ovarian Cancer (n=91) IGS 4 LGS 8 Endometrioid 4 Clear Cell 2 Mucinous	I. Discovery & Technical Validation Archived fresh frozen OC tissue, Prospective benign FFPE fallopian tube tissue Ovarian Cancer (n=57) Benign (n=29) 14 FTE 19 Buffy Coat 14 FTE 19 Buffy Coat



BIOLOGICAL TISSUE VALIDATION – 33 MDMS

Characteristic	Ovarian Cancer (n=105)	Benign Fallopian Tube Controls (n=31)
Age, years (median [IQR])	60 [54-68]	53 [45-62]
BMI, kg/m2 (median [IQR])	28 [23.2-32.9]	26.7 [24.3-33.2]
Pregnancies (median [IQR])	2 [0-3]	3 [2-4]
Live births (median [IQR])	2 [0-3]	2 [1-3]
Race		
White	95 (90%)	27 (87%)
Non-White	5 (5%)	3 (10%)
Unknown	5 (5%)	19 (3%)
Tobacco Use		
Current	11 (10%)	2 (6%)
Previous	28 (27%)	7 (23%)
Never	62 (59%)	22 (71%)
Unknown	4 (4%)	0 (0%)
Menopausal Status		
Premenopausal	13 (12%)	19 (61%)
Perimenopausal	6 (6%)	0 (0%)
Postmenopausal	77 (73%)	12 (39%)
Unknown	9 (9%)	0 (0%)
Histology		
High grade serous	34 (32%)	-
FIGO grade 1/2 endometrioid	21 (20%)	-
FIGO grade 3 endometrioid	7 (7%)	
Clear cell	25 (24%)	-
Well differentiated mucinous	16 (15%)	-
Low grade serous	2 (2%)	
Stage		
I	50 (48%)	-
II	7 (7%)	-
III	39 (37%)	-
IV	9 (9%)	-

OVARIAN CANCER PLASMA CLINICAL PILOT: 11 MDMS

Characteristic	Ovarian cancer (n = 91)	Healthy controls $(n = 91)$
Age, years (median [IOR])	61 [57-68]	61 [58-66]
BML kg/m ² (median [IOR])	27.25 [24.66-30.5]	27.64 [22.95-31.45]
Pregnancies (median [IOR])	2[1-3]	2[1-4]
Live births (median [IOR])	2 [1-3]	211-31
Race	- 10 - 1	-11
White	75 (83%)	89 (98%)
Non-White	13 (14%)	1(1%)
Unknown	3 (3%)	1(1%)
Tobacco Use	- And	
Current	9(10%)	13 (14%)
Previous	23 (25%)	26 (29%)
Never	59 (65%)	52 (57%)
Menopausal Status	C. Starter	
Premenopausal	7 (8%)	9 (10%)
Perimenopausal	5 (5%)	5 (5%)
Postmenopausal	74 (81%)	77 (85%)
Unknown	5 (5%)	0(0%)
Histology		
High grade serous	73 (80%)	
FIGO grade 1 or 2 endometrioid	8 (9%)	-
Clear cell	4 (4%)	S
Low grade serous	4 (4%)	
Mucinous	2 (2%)	-
Stage	1 A	
1	10 (11%)	
II	5 (5%)	-
10	64 (70%)	Sec
IV	12 (13%)	
CA-125 U/mL (Median [IQR])	358.4 [119.2-1044.8]	8.6 [5.8-12.5]





OC histology	Serous	Clear cell	Endometrioid	Mucinous	Mixed
N	76	4	8	2	1
Sensitivity at 95% specificity % (95% CI)	83% (73 - 90%)	75% (19 - 99%)	50% (16 - 84%)	50% (13 - 99%)	100% (3 - 100%)

SOME IMPORTANT CONSIDERATIONS

- Experiments in biospecimen compartment \rightarrow Supernatant, pellet, or homogenate?
- Experiments in stability in at-home collections
 - How many days does the DNA remain stable?
 - Impact of variation in shipping temperatures → Ambient environmental temperature in summer, winter extremes?
 - "People are human" \rightarrow variability inherent in self-collection
- Marker selection refinement \rightarrow MDM down-selection
- Independent validation of MDM panel
- Potential complementary molecular markers
- Voice of the customers \rightarrow patients' and providers' preferences
- Optimizing diversity of at-risk patients \rightarrow Generalizability of the test

NCT05051722: MAYO CLINIC'S ECHO STUDY

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Clinicaltrials.gov

CELEBRATING AN INCREDIBLE TEAM!

Molecular Cancer **Diagnostic Laboratory (Kisiel Lab)** John Kisiel, MD William Taylor Seth Slettedahl Doug Mahoney Kelli Burger **OB/GYN** Research Calise Berger Maureen Lemens, MSN, RN Patrick Foote Karen Ishitani, RN Karen Doering Ann VanOosten Anna Gonser Jainnee Sacksith Collin Chalmers Iris Lin Taylor Rasmusson Stacy Wroblewski Shridhar Lab Viji Shridhar, PhD Julie Staub

External Collaborating Institutions

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and Pathology

Sarah Kerr, MD

Amy Clayton, MD

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THANK YOU!

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